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# **The response of stocks of C, N and P to plant invasion in the coastal wetlands of China**

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24 **Abstract**

25 The increasing success of invasive plant species in wetland areas can threaten their capacity  
26 to store carbon, nitrogen, and phosphorus (C, N, and P). Here we have investigated the  
27 relationships between the different stocks of soil organic carbon (SOC), and total C, N, and  
28 P pools in the plant-soil system from eight different wetland areas across the South-East coast  
29 of China, where the invasive tallgrass *Spartina alterniflora* has replaced the native tall  
30 grasses *Phragmites australis* and the mangrove communities, originally dominated by the  
31 native species *Kandelia obovata* and *Avicennia marina*. The invasive success of *Spartina*  
32 *alterniflora* replacing *Phragmites australis* did not greatly influence soil traits, biomass  
33 accumulation or plant-soil C and N storing capacity. However, the resulting higher ability to  
34 store P in both soil and standing plant biomass (approximately more than 70 and 15 kg P by  
35 ha, respectively) in the invasive than in the native tall grass communities suggesting the  
36 possibility of a decrease in the ecosystem N:P ratio with future consequences to below- and  
37 above-ground trophic chains. The results also showed that a future advance in the native  
38 mangrove replacement by *Spartina alterniflora* could constitute a serious environmental  
39 problem. This includes enrichment of sand in the soil, with the consequent loss of nutrient  
40 retention capacity, as well as a sharp decrease of the stocks of C (2.6 and 2.2 t C ha<sup>-1</sup> in soil  
41 and stand biomass, respectively), N, and P in the plant-soil system. This should be associated  
42 with a worsening of the water quality by aggravating potential eutrophication processes.  
43 Moreover, the loss of carbon and nutrient decreases the potential overall fertility of the system,  
44 strongly hampering the re-establishment of woody mangrove communities in the future.

45

46 **KEYWORDS**

47 Active carbon, plant invasion, soil organic carbon, nutrient stoichiometry

## 48 INTRODUCTION

49 The world contains an estimated 2011 Pg organic carbon of soil (IPCC, 2000). The  
50 importance of soil as a carbon sink is, therefore, crucial for the ability of the Earth to buffer  
51 the increasing levels of atmospheric CO<sub>2</sub> as a consequence of human activities (Joiner et al.,  
52 1999; Smith, 2004; Bonan & Van Cleve, 2011). Wetlands are one of the most sensitive  
53 ecosystems to global climate change. Despite accounting just for 4 to 6% of the land area  
54 (Howe et al., 2009), wetlands accumulate between 20 to 30% of the whole terrestrial soil C  
55 stock (Smith et al., 2004), thus playing a disproportionate role in the global C cycle. As the  
56 ecosystems with the world's highest productivity per unit of area, wetlands combine the dual  
57 function of being a "C source" and a "C sink" (Andreetta et al., 2016). Therefore, even minor  
58 changes in the dynamics of wetland areas will affect global greenhouse gas emissions, which  
59 in turn will influence global warming (Tian et al., 2010). Among wetland areas, estuarine  
60 wetland soils, due to their location at the river-ocean interface, have a particularly large  
61 potential to act as sediment sinks, thus accumulating C (Bianchi et al., 2013; Burden et al.,  
62 2013; Mitsch et al., 2013). In this context, the changes occurred in the last decades in China  
63 coastal wetlands, such as the reduction in area because of sea level rise (Wang et al., 2015b)  
64 and land use change (Wang et al., 2014) are especially important. Despite this reduction,  
65 coastal wetlands in China currently still cover  $5.80 \times 10^4$  km<sup>2</sup> (Wetland China, 2014) and  
66 provide many ecosystem services and products (Liu et al., 2006; Wang et al., 2015a). We  
67 know that increases in tropical storms and ocean levels have altered flooding intensities,  
68 which together with increased pollution and nutrient loads have impacted sedimentary  
69 processes and the capacity of wetlands to store and release C and nutrients (Schewe et al.,  
70 2011; Mendelsohn et al., 2012; Sardans et al., 2012; Ramsar, 2013; Piecuch & Ponte, 2014;

71 Sardans & Peñuelas, 2014). In this context, the success of invasive plant species can also  
72 have an impact on the capacity of wetlands to store and release C, N and P (Wang et al., 2014;  
73 Wang et al., 2015b).

74 Invasive plant species can change the capacity to accumulate N and P in biomass and  
75 soil (Wang et al., 2015a). The impacts of successful invasive plants on the concentrations and  
76 stoichiometry of soil nutrients have been widely studied (Funk & Vitousek, 2007; Sardans &  
77 Peñuelas, 2012), and some general trends have emerged (González et al., 2010; Matzek, 2011;  
78 Sardans & Peñuelas, 2012; Sardans et al., 2017). The impacts of invasive plants can differ  
79 and even be opposite depending on the natural availability levels of soil nutrients and,  
80 generally, on the capacity of a site to sustain low or high plant production in accordance with  
81 the soil and climatic conditions (González et al., 2010; Matzek, 2011; Sardans & Peñuelas,  
82 2012; Sardans et al., 2017).

83 The active C from the total soil organic carbon, is highly susceptible to oxidation and  
84 decomposition and is strongly influenced by plants and microorganisms (Kimura et al., 2004;  
85 Chen et al., 2010) and includes dissolved organic carbon (DOC), labile organic carbon (LOC),  
86 and soil microbial biomass organic carbon (MBC). Differently, the total content of soil  
87 organic carbon (SOC) changes over a long-time scale, and their fluctuations are not easily  
88 discerned within a short time period (Wissing et al., 2011). Therefore, distinguishing between  
89 the active-C fraction from the total SOC pool is important to assess the effect of plant  
90 invasion on soil C dynamics. Different forms of active SOC have different sensitivities to  
91 environmental change (Gu et al., 2004), but few studies have examined the relationships  
92 between the different forms of active SOC and changes in other environmental parameters  
93 (Xu et al., 2011; Zhao et al., 2014), and especially of soil properties. Active SOC is a major  
94 source of CO<sub>2</sub> and CH<sub>4</sub> produced by microbes, so properly managing this pool of active C is

95 important for mitigating global climate change (Knoblauch et al., 2011; Hanke et al., 2013).  
96 This C:nutrient stoichiometry is a good indicator of changes in soil C dynamics in wetlands  
97 due to changes in the environment, thereby providing information of the impacts on nutrient  
98 cycling and status and, thus, is informative about the potential capacity of these ecosystems  
99 to fix C and reduce the impact of the emissions of C-source greenhouse gases (Peñuelas et  
100 al., 2012 and 2013).

101         The impacts of plant invasion on total soil C accumulation in the various soil fractions  
102 and the further relationships of these impacts on C with other soil traits, such as C:N:P ratios,  
103 are also unresolved (Sardans & Peñuelas, 2012). Most available data generally suggest that  
104 the low costs of foliar construction and high phenotypic plasticity in taking up available  
105 nutrients frequently contribute to invasive species success in nutrient-rich environments  
106 (Daehler, 2003; González et al., 2010; Sardans & Peñuelas, 2012), whereas in nutrient-poor  
107 soils the success of invasive plants would depend on more conservative strategies, such as a  
108 higher nutrient-use efficiency (Funk & Vitousek, 2007; González et al., 2010; Matzek, 2011;  
109 Sardans et al., 2017), especially on short timescales (Funk & Vitousek, 2007). The existence  
110 of numerous exceptions, however, prevent this question from being totally resolved (Sardans  
111 & Peñuelas, 2012).

112         Some studies have discussed soil C dynamics and nutrient stoichiometry (Tian et al.,  
113 2010; Schipper & Sparling, 2011), but few have distinguished them at a fine scale in soil  
114 profiles. Studies addressing the relationships of species invasive success with soil C and  
115 nutrient status has been mainly focused on the first centimeters of soil (Sardans & Peñuelas,  
116 2012). However, there is little available information about the impact of plant invasive  
117 success on soil depth. The different soil textures and soil organic matter decomposition  
118 observed in superficial soil layers linked to plant alien success, can thereafter produce a shift

119 in the accumulation of some organic carbon fractions along soil deep layers (Senga et al.,  
120 2011; Xiang et al., 2015). The impacts of plant invasion on the distribution of total soil C  
121 accumulated among the various soil fractions and the further relationships of these impacts  
122 on C dynamics with other soil traits, such as C:N:P ratios, are also unclear (Sardans &  
123 Peñuelas, 2012). We hypothesized that changes in soil C storage along the soil profile could  
124 be related to different plant species dominating above-ground. The knowledge of the  
125 variation in C accumulation along the vertical soil profile and its causes is crucial for  
126 understanding the capacity of wetlands to act as carbon sinks in absolute amounts (Craft,  
127 2007).

128 We aimed (i) to determine whether the success of plant invasion of the same invasive  
129 species at different sites was based on a similar strategy of C, N, and P use, and (ii) to compare  
130 the differences of soil traits and plant-soil system C, N, and P stocks between successful  
131 recently established invasive communities of *Spartina alterniflora* in two very different  
132 native communities: native tallgrass and mangrove communities with respect pure non-  
133 invaded native communities in different areas along subtropical and tropical coastal wetlands  
134 of China.

135

## 136 **MATERIALS AND METHODS**

### 137 **Study area**

138 We studied the coastal wetlands of central-south China, where today plant invasion effect is  
139 one of the most important problems (Figure 1). In 1979, *Spartina alterniflora* was introduced  
140 into China from the United States for the purpose of accumulating silt, thereby protecting  
141 beaches and berms. However, *S. alterniflora* was identified as an efficient invasive plant,

142 decreasing the area of indigenous vegetation communities, transforming the habitat, and  
143 jeopardizing the survival of the animal species in the intertidal zone, and ultimately  
144 producing a series of ecological and economic hazards to the area (Yang et al., 2009). *S.*  
145 *alterniflora* is the only coastal wetland invasive plant identified by the State Environmental  
146 Protection Administration of China since 2003. *S. alterniflora* has an apparently superior  
147 breeding ability and adaptability than several native wetland species. Thus, it has spread  
148 wildly and invaded and replaced coastal native plant communities where it forms mono-  
149 species communities. Today, *S. alterniflora* grows from the North of China from the Yalu  
150 River estuary to the South in Guangxi Beibu Gulf. The distribution area accounts for about  
151 2% of the wetland area in China (Lu & Zhang, 2013). China is the country with the largest  
152 outbreak of *S. alterniflora* in the world, especially in its subtropical coastal wetland. The  
153 distribution area of *S. alterniflora* reaches more than 92.44% of the total invaded areas in the  
154 subtropical coastal wetland (Lu & Zhang, 2013). Therefore, the subtropical coastal wetland  
155 is an ideal area for the study of *S. alterniflora* invasion.

156

### 157 **Sample collection and measurements**

158 Soil samples were collected during the plants' main growing season (June to August) in 2015  
159 from eight locations in China's subtropical and tropical coastal wetlands (Figure 1). We  
160 sampled soils in well established *Spartina alterniflora* communities in sites originally  
161 dominated by *Phragmite australis* (grass), *Kandelia obovata* (mangrove), *Avicennia marina*  
162 (mangrove), *Spartina alterniflora* (invaded from 7 to 15 years ago), and also collected soils  
163 from native plants communities growing at these sites for more than 30 years. In our study,  
164 for the *S. alterniflora*-invaded *Phragmite australis* site, the native plants were completely  
165 replaced by *S. alterniflora*. However, for the *S. alterniflora*-invaded mangrove site, the



166 invasion has begun in mangrove community margins and empty spaces within the mangrove  
167 community, and it is currently spreading its cover by all the mangrove community space.  
168 Three plots were randomly established in each community type, and soil profiles (width, 1  
169 m; length, 0.6 m; depth, 0.4 m) were excavated. The samples were collected with a small  
170 sampler (length, 0.4 m; diameter, 0.1 m) from each of four soil layers (0–10, 10–20, 20–30,  
171 and 30–40 cm) at the center and both ends of the soil pits. These three samples from each  
172 layer were bulked to form one composite sample per layer. A total of 240 soil samples (ten  
173 sampling site  $\times$  two communities  $\times$  three plots  $\times$  four soil layers) were thus collected. The  
174 core samples were divided into two parts, with one part unprocessed for the measurement of  
175 soil MBC and DOC, and the other part air-dried and finely grounded in a ball mill after the  
176 removal of all roots, and visible plant remains used for the determination of total SOC and  
177 LOC. Total SOC was measured with a Vario EL III Elemental Analyzer (Elementar Scientific  
178 Instruments, Hanau, Germany), DOC by extracting the soils with deionized water (1:5 ratio)  
179 and measuring the C concentration using a TOC-V CPH total C analyzer (Shimadzu  
180 Scientific Instruments, Kyoto, Japan), MBC by fumigation-extraction (Lu, 1999), and LOC  
181 by digestion with 333 mM KMnO<sub>4</sub> (Xu et al., 2011; Wang et al., 2015c).

182 Bulk density was measured for the three bulked cores (5 cm diameter, 3 cm depth)  
183 collected from each soil layer. Soil salinity, pH, and particle-size distribution were measured  
184 by a DDS-307 salinity meter (Boqu Scientific Instruments, Shanghai, China), an 868 pH  
185 meter (Orion Scientific Instruments, Minnesota, USA), and a Master Sizer 2000 Laser  
186 Particle Size Analyzer (Malvern Scientific Instruments, Suffolk, UK), respectively. Soil total  
187 N concentration was determined with a Vario EL III Elemental Analyzer (Elementar  
188 Scientific Instruments, Hanau, Germany), and total soil P concentration was determined by  
189 perchloric-acid digestion followed by ammonium-molybdate colorimetry and measurement

190 using a UV-2450 spectrophotometer (Shimadzu Scientific Instruments, Kyoto, Japan).

191 Tallgrass above-ground biomass in each plot was collected from a randomly selected  
192 quadrat (10 × 10 m), and the above-ground biomasses were collected from a selected center  
193 sub-quadrat (1 × 1 m). For the mangrove, we only collected some above-ground materials,  
194 not all of the above-ground biomass. At the same time, we determined the mangrove diameter  
195 at the breast height and the height and density of all plants. Also, the above-ground mangrove  
196 biomass of each plant population was calculated by a previously reported equation (Tam et  
197 al., 1995).

198 All plant material was gently washed with deionized water and then oven-dried to a  
199 constant mass (80 °C for 24–36 h) and weighed. The concentrations of C and N of the plants  
200 were determined using a Vario EL III Elemental Analyzer (Elementar Scientific Instruments,  
201 Hanau, Germany). Total plant P concentrations were all determined by the method of  
202 colorimetrically using a UV-2450 spectrophotometer (Shimadzu Scientific Instruments,  
203 Kyoto, Japan) at a wavelength of 700 nm (Lu, 1999). C, N, and P stocks in the plant were  
204 calculated by the biomass multiplied by the nutrient concentration.

205

## 206 **Statistical analyses**

207 We performed general mixed models (GLM) with plant type (invasive versus native) as an  
208 independent fixed categorical variable, site as an independent random factor, and soil traits  
209 plant above-ground biomass, and C, N, and P stocks in plant above-ground biomass as  
210 continuous dependent variables. We used the “*nlme*” (Pinheiro et al., 2016) R package with  
211 the “*lme*” function. If the variable was not normally distributed, it was log-transformed. We  
212 chose the best model for each dependent variable based on the Akaike information criterion.

213 We used the MuMIn (Barton, 2012) R package in the mixed models to estimate the  
214 percentage of the variance explained by the mixed models. We conducted Tukey's post hoc  
215 tests to detect significant differences in the analyses for more than two communities using  
216 the "*multcomp*" (Hothorn et al., 2013) R package with the "*glht*" function.

217 We also performed other multivariate statistical analyses. We determined the overall  
218 differences in the changes of the soil C fractions, N, and P concentrations, stoichiometric  
219 ratios, salinity, pH, and temperature in the species-specific plant communities using general  
220 discriminant analysis (GDA), including the component of the variance due to the different  
221 soil layers as an independent categorical variable. Discriminant analyses consist of a  
222 supervised statistical algorithm that derives an optimal separation between groups established  
223 a priori by maximizing between-group variance while minimizing within-group variance  
224 (Raamsdonk et al., 2001). GDA is thus an adequate tool for identifying the variables most  
225 responsible for the differences among groups, while controlling the component of the  
226 variance due to other categorical variables. The GDAs were performed using Statistica 6.0  
227 (StatSoft, Inc. Tulsa, USA).

228

## 229 **RESULTS**

### 230 ***Spartina alterniflora* invasion of tallgrass wetlands**

231 Compared to the original native communities of *Phragmite australis*, the *Spartina*  
232 *alterniflora* communities that currently occupy wetland areas previously inhabited by the  
233 native tallgrass *P. australis* had higher soil C in microbial biomass, total soil C and P content,  
234 soil C:N ratio, and P content in above-ground biomass, but lower soil N:P ratio and C stocks  
235 in above-ground plant biomass (Table 1, Figures 2 and 3). Our data analyses provide  
236 evidences of higher soil C (approx. 18.8 versus 17.5 t C ha<sup>-1</sup>) in the invaded than native

237 community but smaller C stocks in aboveground biomass (approx. 6 versus 8 t C ha<sup>-1</sup>) in the  
238 invaded than in native communities, thus resulting in a trend to less total C stored in the plant-  
239 soil system, irrespective of the C stored in below-ground biomass. Additionally, our data  
240 indicate higher amounts of P stored in soil (approx. 0.07 t P ha<sup>-1</sup>) and in aboveground biomass  
241 (approx. 1.5 g m<sup>-2</sup>, that is 0.015 t P ha<sup>-1</sup>) in invasive than in native communities. Therefore,  
242 without knowing the P stored in the belowground biomass, the data showed a total higher P  
243 stored in the invasive than native communities.

244 The GDA analysis with soil data, comparing the communities dominated by  
245 *Phragmites australis* with those dominated by *Spartina alterniflora*, showed no differences  
246 in main soil features (Squared Mahalanobis = 2.46, F = 1.01, P = 0.46) between the  
247 *Phragmites australis* and *Spartina alterniflora* communities.

248

#### 249 ***Spartina alterniflora* invasion of mangrove**

250 In our study, the *Spartina alterniflora* communities that currently occupy areas in the margins  
251 of mangrove community and in the empty spaces within the mangrove community, which  
252 previously were occupied by native mangrove communities (mostly previously inhabited by  
253 *Kandelia obovata* or *Avicennia marina*), had lower soil organic carbon, total soil C and P  
254 content, and soil C:N, C:P, and N:P ratios (Table 2). Moreover, *Spartina alterniflora*  
255 communities also had lower C, N, and P stocks in the above-ground plant biomass (only  
256 compared with *Avicennia*-dominated mangroves) and lower aboveground biomass than  
257 native mangrove communities (Figures 2 and 3). Hence, data analyses indicate higher stocks  
258 of soil C (approx. 2.6 t C ha<sup>-1</sup>), N (approx. 0.17 t N ha<sup>-1</sup>), and P (approx. 0.05 t P ha<sup>-1</sup>) and  
259 also higher stocks in the above-ground biomass of C (approx. 2.2 t C ha<sup>-1</sup>), N (approx. 0.65 t  
260 N ha<sup>-1</sup>), and P (approx. 0.08 t P ha<sup>-1</sup>) in the native than invaded community (Figures 2 and 3).

261 Thus resulting in a trend to substantially less C, N, and P stored in the invaded plant-soil  
262 system relative to the native-dominated mangrove communities, irrespective of the C, N, and  
263 P stored in below-ground biomass.

264 In contrast, in this case, the GDA analysis of the soil data, comparing the *Spartina*-  
265 dominated with the native mangrove communities, revealed significant differences (Squared  
266 Mahalanobis = 9.84,  $F = 17.3$ ,  $P < 0.0001$ ) arising after the invasion (Figure 4).

267

## 268 **DISCUSSION**

### 269 **General descriptive results**

270 We have not observed important changes in C:N and C:P soil ratios along the soil profile.  
271 This is an interesting and particular result of these studied wetland soils, which is probably  
272 related to the sediment loadings that these soils continuously receive and the anoxic  
273 conditions that prevent a fast and complete decomposition of the litter, which is incessantly  
274 covered by new sediments and progressively buried to deeper soil layers. A trend to uniform  
275 organic carbon contents along the vertical soil profile in wetlands has been previously  
276 observed in soils receiving regular loads of sediments (Alongi et al., 2001; Senga et al., 2011),  
277 which is related to the physical protection of soil organic carbon linked to sedimentation  
278 typical from seasonally-flooded wetlands (Maynard et al., 2011).

279

### 280 **The relationships between species invasion in tallgrass wetland communities and soil** 281 **traits and plant-soil C, N, and P stocks**

282 Our results show that the invasion of *Spartina alterniflora* replacing *Phragmites australis*,  
283 when comparing adult communities of both species, had no clear impacts on the overall C  
284 storing capacity of the plant-soil systems. Although the invaded community had more C in

285 the soil, there was a correspondingly lower amount of C stored in the above-ground biomass.  
286 Previous studies conducted elsewhere in Chinese wetlands and investigating the impact of *S.*  
287 *alterniflora* on soil properties after replacing tallgrass communities dominated by other  
288 native species, such as *Cyperus malaccensis* or *Suaeda salsa*, also showed a trend to  
289 increasing soil C stocks with time since invasion (Chen et al., 2015; Jin et al., 2017; Liu et  
290 al., 2017). In light of these previous results, our data appear consistent with the general view  
291 that invasive processes in wetlands tend to maintain or even increase C stocks in the soil, as  
292 observed at the global level (Liao et al., 2008).

293 Reports addressing the impacts of *S. alterniflora* on soil stocks of N and P and on the  
294 stocks of C, N, and P in plant biomass are rare. After the replacement of *P. australis* by *S.*  
295 *alterniflora*, we found a general increase in P stocks both in the soil and above-ground  
296 biomass, but not in N, thus resulting in a decrease in soil and plant N:P ratios. To our  
297 knowledge, so far there are very few equivalent studies with which we can compare our  
298 results. Likewise, Wang et al. (2015a) found a decrease in N:P ratios in a wetland area of the  
299 Minjiang River, where *S. alterniflora* was invading the native *Cyperus malaccensis* tall grass  
300 communities, although they did not observe any effects of invasion on soil P concentrations.

301 The higher P contents in soil observed after the invasion was mainly due to the higher  
302 bulk density in the invaded than in the native communities. Bulk density has been positively  
303 correlated with organic C, N, and P concentrations in similar wetland areas in the soils below  
304 *Carex* sp. and *Phragmites australis* (Peng et al., 2005). Differences and fluctuations in bulk  
305 density have also been frequently associated with the success of invasive plant species (Miller  
306 et al., 2006; Lortie & Cushman, 2007; Pande et al., 2007). Summarizing, the successful  
307 invasion of *Spartina alterniflora* replacing *Phragmites australis* in Chinese wetlands should  
308 not produce great impacts on their C storing capacity, but it could increase soil and ecosystem

309 P-storing ability due to the higher plant P concentrations and also due to the increase in soil  
310 bulk density, further decreasing the N:P ratios at an ecosystem level with likely consequences  
311 for the whole soil and above-ground trophic chains.

312

313 **The relationships between species invasion in mangrove communities and soil traits and**  
314 **plant-soil C, N, and P stocks**

315 The lower soil bulk densities in *S. alterniflora*-invaded communities relative to the native  
316 mangrove communities were mainly related to a lower sand content in the latter. The success  
317 of *S. alterniflora* was thus associated with decreases in bulk density and total soil C, N, and  
318 P contents per unit of surface area.

319 As in other studies that have observed differences in bulk density and elemental  
320 composition between soils in invaded and native communities (Wang et al., 2018), the causes  
321 of these differences in our study were unclear. Nevertheless, this result suggests that the  
322 scarcer root system of a tallgrass community, compared with the more consistent root system  
323 of a mangrove stand, could have a lower ability to retain small-size particles in the soil, such  
324 as clay.

325 In this case, the large differences in above-ground biomass between the native  
326 mangrove stands compared to the *Spartina alterniflora*-dominated tallgrass community can  
327 easily explain the great losses in C, N, and P stocks in the soils after the invasion. Changes  
328 in soil traits due to plant invasion, such as SOC concentration (Zhou et al., 2015), texture  
329 (Uselman et al., 2014; Haubensak et al., 2014), and other soil parameters (Blank et al., 2015),  
330 have been correlated with differences in plant or root biomass and/or shoot:root ratio between  
331 native and invaded plant communities. In fact, soil properties such as texture can change in  
332 wetlands within 10 years, as observed in other studies (Craft et al., 1999 and 2002). Other

333 works have also observed changes in soil bulk density (Zhang et al., 2009), C contents  
334 (Koutika et al., 2007; Koteen et al., 2011; Throop et al., 2013; Yu et al., 2014), and several  
335 other traits (Yu et al., 2014; Souza-Alonso et al., 2015) as a consequence of successful plant  
336 invasion. The similar SOC values along the vertical soil profile shown earlier suggest that  
337 these wetlands are sinks of sediments, reinforcing the idea that sites with different vegetation  
338 cover with different structure and root system size can change soil structure by favoring  
339 sedimentation of different particle size classes. Thus, our study is consistent with the  
340 observation that soil texture is a sensitive factor during plant invasion, as has been observed  
341 in several previous studies (Craft et al., 1999 and 2002; Surrrette & Brewer, 2008; Zhang et  
342 al., 2009). The evidence emerging from many previous investigations generally indicates that  
343 the differences in soil properties between invaded and original native communities are  
344 associated with the distinct structural and functional traits of the invasive versus the native  
345 species; namely nutrient uptake, root system size and structure, litter production, foliar  
346 nutrient resorption, and growth rate (Lindsay & French, 2005; Titus, 2009; Sardans &  
347 Peñuelas, 2012; Novoa et al., 2014).

348         Nevertheless, the replacement of mangrove woody ecosystems by tallgrass  
349 formations is clearly associated with an extensive change in the physical and chemical traits  
350 of the whole ecosystem, with biomass loss, reductions in soil bulk density, and depletions of  
351 C, N, and P stocks in the plant-soil system. These findings suggest that an eventual future  
352 regeneration process would be difficult because the ecosystem has become a lot less fertile  
353 and with sandy soil, conditions that are unfavorable for the reestablishment of the original  
354 woody community, whose higher biomass requires greater resources that largely exceeds the  
355 supply capacity of the current invaded system. Additionally, the lower ability to retain N and  
356 P has negative consequences for water quality since it promotes eutrophication processes.



357 Moreover, these elements combined also worsen the overall C and nutrients storing capacity  
358 of these wetlands and should be considered as a general process reducing the environmental  
359 quality of the ecosystem.

360

## 361 **FINAL REMARKS AND CONCLUSION**

362 We show that the strong ecosystem impacts on soil characteristics, biomass accumulation,  
363 and plant-soil system C, N, and P stocks after the successful invasion of *S. alterniflora* are  
364 clearly different depending on the original native community being replaced.

365         When replacing the native grass *P. australis*, *S. alterniflora* did not produce a change  
366 in soil traits, biomass accumulation or plant-soil C and N storing capacity. However, a higher  
367 ability to store more P in both the soil and stand plant biomass suggests the possibility of a  
368 decrease in ecosystem N:P ratios, which may imply future consequences for the whole below-  
369 and above-ground trophic chains. Secondly, a higher efficiency to absorb P might improve  
370 the P-filtering performance by the ecosystem, thus better preventing the eutrophication of  
371 water.

372         However, the replacement of the native mangrove communities by *S. alterniflora*  
373 resulted in several critical impacts and involved a serious environmental problem. These  
374 included an enrichment of sand in soil with a consequent loss of nutrient retention capacity,  
375 the loss of great amounts of C, N, and P out of the plant-soil system with a general loss of  
376 plant production and decay of water quality, thereby exacerbating potential eutrophication  
377 processes. Moreover, the loss of C and nutrient decreased the overall fertility of the system,  
378 strongly hampering the future re-establishment of the original woody mangrove communities.

379

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389

#### 390 **CONFLICTS OF INTEREST**

391 The authors declare no conflicts of interest.

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658 **Figure Captions**

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660 **FIGURE 1** The location of the study area and sampling sites in China.

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662 **FIGURE 2** C, N, and P stocks (mean  $\pm$  S.E.) in above-ground biomass of invasive tallgrass  
663 wetland communities of *Spartina alterniflora* (in black) replacing tall grass native  
664 *Phragmites australis* community wetlands (in red) and also replacing mangrove (dominated  
665 by the native species *Avicennia marina*) communities (in green).

666

667 **FIGURE 3** Above-ground biomass (mean  $\pm$  S.E.) of invasive tallgrass wetland communities  
668 of *Spartina alterniflora* (in black) replacing tall grass native *Phragmites australis* community  
669 wetlands (in red) and replacing mangrove (dominated by the native species *Kandelia obovata*)  
670 communities (in blue) and also replacing mangrove (dominated by the native species  
671 *Avicennia marina*) communities (in green).

672

673 **FIGURE 4** Results of the general discriminant analysis (GDA) with the different studied soil  
674 variables as independent continuous variables, different sites as categorical controlling  
675 independent variable, and community types (Mangrove and *Spartina alterniflora*  
676 communities) as dependent grouping categorical variable.

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705 **TABLE 1** Results of the mixed models of sites where *Spartina alterniflora* has replaced  
706 native *Phragmites australis* tallgrass with different studied plant and soil variables as  
707 dependent factors, species invasion as fixed factor, and site and soil depth as random values  
708 in the case of soil variables. SOC = soil organic carbon, DOC = dissolved organic carbon,  
709 LOC = labile organic carbon, MBC = microbial biomass organic carbon. Significant  
710 responses ( $P < 0.05$ ) to *S. alterniflora* invasion is highlighted in bold type

Dependent variables	Mixed model					
	lme(Dependent variable ~ invasion, data = daDES, random = list(~1 site, ~1 soildepth))					
	Fixed Factor (invasion)				Model results	
	Native	Invasive	F	P-value	R <sup>2</sup> m	R <sup>2</sup> c
SOC (mg g <sup>-1</sup> )	21.2±0.6	20.9±0.6	0.28	0.60	0.0022	0.48
LOC (mg g <sup>-1</sup> )	3.69±0.24	3.63±0.17	0.055	0.82	0.00051	0.34
DOC (mg g <sup>-1</sup> )	118±8	121±10	1.44	0.23	0.00093	0.95
<b>MBC (mg g<sup>-1</sup>)</b>	<b>300±22</b>	<b>336±24</b>	<b>13.2</b>	<b>&lt;0.0001</b>	0.012	0.94
<b>Soil Total C content (t ha<sup>-1</sup>)</b>	<b>17.5±0.6</b>	<b>18.9±0.7</b>	<b>4.36</b>	<b>0.041</b>	0.024	0.61
Soil Total N content (t ha <sup>-1</sup> )	1.21±0.05	1.22±0.06	0.016	0.90	0.0001	0.57
<b>Soil Total P content (t ha<sup>-1</sup>)</b>	<b>0.783±0.027</b>	<b>0.850±0.033</b>	<b>11.3</b>	<b>0.0014</b>	0.027	0.83
<b>Soil C:N</b>	<b>15.1±0.7</b>	<b>16.4±0.9</b>	<b>21.3</b>	<b>&lt;0.0001</b>	0.014	0.95
Soil C:P	22.4±0.5	22.4±0.5	0.0009	0.98	0.0001	0.17
<b>Soil N:P</b>	<b>1.62±0.09</b>	<b>1.52±0.09</b>	<b>4.12</b>	<b>0.047</b>	0.0072	0.88
<b>Plant height</b>	<b>2.63±0.10</b>	<b>1.60±0.05</b>	<b>130</b>	<b>&lt;0.0001</b>	0.53	0.71
<b>Plant density (plant m<sup>-2</sup>)</b>	<b>90.9±9.2</b>	<b>155±9</b>	<b>220</b>	<b>&lt;0.0001</b>	0.20	0.93
<b>Soil [N] (g kg<sup>-1</sup>)</b>	<b>1.55±0.10</b>	<b>1.43±0.10</b>	<b>4.80</b>	<b>0.033</b>	0.0080	0.88
Soil [P] (g kg <sup>-1</sup> )	0.944±0.013	0.931±0.012	1.99	0.16	0.0064	0.77
Soil pH	7.32±0.19	7.40±0.18	2.77	0.10	0.00089	0.98
Soil salinity (mS cm <sup>-1</sup> )	1.92±0.29	1.97±0.30	0.20	0.65	0.00014	0.95
Soil water content (%)	41.9±1.5	40.8±1.6	2.29	0.14	0.0028	0.91
<b>Soil bulk density (g cm<sup>-3</sup>)</b>	<b>0.851±0.037</b>	<b>0.912±0.039</b>	<b>7.09</b>	<b>0.01</b>	0.014	0.86
Soil Clay content (%)	14.1±0.7	14.6±0.8	0.57	0.45	0.0023	0.71
<b>Soil silt content (%)</b>	<b>65.1±1.3</b>	<b>62.4±1.4</b>	<b>14.8</b>	<b>&lt;0.0001</b>	0.022	0.90
<b>Soil sandy content (%)</b>	<b>20.7±1.8</b>	<b>23.0±2.1</b>	<b>5.37</b>	<b>0.024</b>	0.0071	0.91

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713 **TABLE 2** Results of the mixed models of sites where a *Spartina alterniflora* have replaced  
714 a native mangrove community with different studied plant and soil variables as dependent  
715 factors, species invasion as fixed factor and site and soil depth as random values in the case  
716 of soil variables. SOC = soil organic carbon, DOC = dissolved organic carbon, LOC = labile  
717 organic carbon, MBC = microbial biomass organic carbon. Significant responses ( $P<0.05$ )  
718 to *Spartina alterniflora* invasion is highlighted in bold type  
719

Dependent variables	Mixed model					
	lme(Dependent variable ~ invasion, data = dades, random = list(~1 site, ~1 soildepth))					
	Fixed Factor (invasion)				Model results	
	Native	Invasive	F	P-value	R <sup>2</sup> m	R <sup>2</sup> c
SOC (mg g <sup>-1</sup> )	<b>16.6±0.5a</b>	<b>14.8±0.4b</b>	<b>41.9</b>	<b>&lt;0.0001</b>	0.034	0.86
LOC (mg g <sup>-1</sup> )	4.20±0.15	4.29±0.14	0.067	0.80	0.00033	0.17
DOC (mg g <sup>-1</sup> )	77.3±4.1	74.2±2.6	0.55	0.46	0.0025	0.25
<b>MBC (mg g<sup>-1</sup>)</b>	337±22	305±21	3.2	0.078	0.0060	0.69
<b>Soil Total C content (t ha<sup>-1</sup>)</b>	<b>13.3±0.3</b>	<b>10.9±0.3</b>	<b>89.1</b>	<b>&lt;0.0001</b>	0.16	0.71
Soil Total N content (t ha <sup>-1</sup> )	<b>1.20±0.03</b>	<b>1.03±0.02</b>	<b>45.8</b>	<b>&lt;0.0001</b>	0.10	0.63
<b>Soil Total P content (t ha<sup>-1</sup>)</b>	<b>0.570±0.017</b>	<b>0.526±0.014</b>	<b>10.3</b>	<b>0.0017</b>	0.020	0.68
<b>Soil C:N</b>	<b>11.2±0.2</b>	<b>10.7±0.2</b>	<b>9.16</b>	<b>0.0029</b>	0.016	0.71
Soil C:P	<b>24.6±0.7</b>	<b>21.2±0.5</b>	<b>42.5</b>	<b>&lt;0.0001</b>	0.072	0.72
<b>Soil N:P</b>	<b>2.30±0.10</b>	<b>2.06±0.07</b>	<b>15.2</b>	<b>&lt;0.0001</b>	0.017	0.81
<b>Plant height</b>	<b>3.14±0.19</b>	<b>1.49±0.04</b>	<b>128</b>	<b>&lt;0.0001</b>	0.28	0.64
<b>Plant density (plant m<sup>-2</sup>)</b>	<b>1.29±0.07</b>	<b>100±10</b>	<b>202</b>	<b>&lt;0.0001</b>	0.35	0.71
<b>Soil [N] (g kg<sup>-1</sup>)</b>	<b>1.48±0.05</b>	<b>1.39±0.04</b>	<b>12.6</b>	<b>&lt;0.0001</b>	0.012	0.84
Soil [P] (g kg <sup>-1</sup> )	0.710±0.025	0.723±0.022	1.87	0.17	0.00086	0.92
Soil pH	<b>6.14±0.13</b>	<b>6.65±0.13</b>	<b>24.7</b>	<b>&lt;0.0001</b>	0.035	0.76
Soil salinity (mS cm <sup>-1</sup> )	4.21±0.22	4.29±0.25	0.11	0.74	0.00014	0.80
Soil water content (%)	<b>46.3±0.9</b>	<b>49.6±1.1</b>	<b>13.5</b>	<b>0.0003</b>	0.026	0.68
<b>Soil bulk density (g cm<sup>-3</sup>)</b>	<b>0.855±0.025</b>	<b>0.774±0.026</b>	<b>31.2</b>	<b>&lt;0.0001</b>	0.022	0.88
Soil Clay content (%)	13.8±0.6	14.6±0.5	1.81	0.18	0.0050	0.54
<b>Soil silt content (%)</b>	<b>58.5±2.1</b>	<b>60.5±2.0</b>	<b>5.26</b>	<b>0.023</b>	0.0021	0.93
<b>Soil sandy content (%)</b>	<b>27.6±2.6</b>	<b>24.8±2.4</b>	<b>5.46</b>	<b>0.021</b>	0.0028	0.91

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